

^{68}Ga -citrate PET/CT is Valuable for Abdominal Lymphoma Imaging

Ling Wang, Jiong Cai, Hong Yang, Zhaohui Zhu, Fang Li

Abstract—Introduction: Although ^{67}Ga -citrate has been used in scintigraphic imaging for lymphoma, the long physical half-life of ^{67}Ga and inferior resolution of SPECT scanner limited the wide-spread application of this agent. In contrast, the ideal physical half-life of ^{68}Ga and superior resolution of PET scanner push us to test lymphoma imaging potential of ^{68}Ga -citrate. Materials: Sodium citrate was added into newly eluted ^{68}Ga and incubated at room temperature for 15min for ^{68}Ga -citrate preparation. Three patients with abdominal lymphoma were retrospectively analyzed after ^{68}Ga -citrate PET/CT imaging. Results: ^{68}Ga -citrate was labeled with >99% yield and purity within 15 minutes. ^{68}Ga -citrate PET/CT revealed rapid progression of bowel diffuse large B-cell lymphoma in one patient; ^{68}Ga -citrate revealed more lesion than ^{18}F -FDG in another diffuse large B-cell lymphoma patient; ^{68}Ga -citrate imaging was negative in an enteropathy-associated T cell lymphoma patient. Conclusion: A simplified method to prepare ^{68}Ga -citrate without nuclide pre-purification is available. ^{68}Ga -citrate can be used for lymphoma imaging and further evaluation is warranted.

Index Terms— ^{68}Ga -Citrate, Diffuse large B-cell Lymphoma, PET, imaging.

INTRODUCTION

Lymphoma is a type of cancer that occurs when B or T lymphocytes divide faster or live longer than they should be. Lymphoma often originate in lymph nodes and may develop in the lymph nodes, spleen, bone marrow, blood or extranodal sites including tonsils, skin, brain, bowels and bone and eventually they form a tumor [1, 2]. Lymphoma presents with certain non-specific symptoms such as swelling of lymph nodes, fever, night sweats, weight loss, anorexia, fatigue, dyspnea, itching, and etc. Lymphoma is definitively diagnosed by a lymph node biopsy, further characterized by immunophenotyping, flow cytometry, FISH testing, and finally divided into different categories according to whether or not it is a Hodgkin lymphoma, the site that the cell arises from, whether the cell that is replicating is a T cell or B cell or natural killer cell [3,4]. Indolent lymphoma is compatible with a long life even without treatment, whereas

aggressive lymphoma causes rapid deterioration and death [5]. However, Most of the aggressive lymphomas such as diffuse large B-cell lymphoma (DLBCL) and enteropathy-associated T-cell lymphoma (EATL) respond well to treatment and are curable [6]. The correct diagnosis and classification of the disease is very important to the prognosis. DLBCL is aggressive lymphoma often occurred outside lymph nodes in all ages, but most commonly in older adults with overall 5-year survival of 60%. Its relative incidence in adults is 40 to 50% of lymphomas. Most DLBCL resemble B cells of large germinal centers and express CD20 on cell surface [7]. EATL, a peripheral T-cell lymphoma, accounted for 9% of gastrointestinal lymphomas [8].

^{67}Ga -citrate has been used in scintigraphic imaging for lymphoma and was valuable for prognosis [9-12]. ^{67}Ga scintigraphy is an excellent predictor of residual tumor viability in DLBCL patients and that persistent positivity of the scan predicts poor outcome and may justify a change in treatment [13]. However, the long physical half-life of ^{67}Ga (78h) limited the wide-spread application of this agent because the injection dose is relatively low, which result in low image quality. The gamma spectrum of ^{67}Ga varies from 92 to 300keV, which lower the quality and resolution of the final image and increase the radiation dose to patient and clinical staff. In recent years, ^{68}Ga has been produced successfully and is commercially available. The ideal physical half-life of ^{68}Ga (68min) makes ^{68}Ga -citrate can be administered with high dose. The positron emitted by ^{68}Ga can be used for PET imaging with high resolution and high image quality. Interestingly, ^{68}Ga -citrate has been successfully in the diagnosis of osteomyelitis, diskitis and intra-abdominal infection but not in lymphoma [14, 15]. Also, the relatively complex procedure of ^{68}Ga -citrate preparation in the literature makes the routine use of ^{68}Ga -citrate difficult [14, 16]. In this study, we try a simplified method to label citrate with ^{68}Ga and test its imaging potential in lymphoma patients.

EXPERIMENTAL SECTION

Materials

$^{68}\text{Ge}/^{68}\text{Ga}$ generator was provided by ITG (Isotope Technologies Garching GmbH, Germany). ^{68}Ga was eluted with 0.05N HCl from the generator. Sodium citrate was produced by Xinning pharmaceutical (Guangdong, China).

^{68}Ga -citrate labeling and quality control

Sodium citrate of 0.1mol/L was prepared with Millipore water (18M Ω). Aliquoting 0.2ml citrate into 0.6ml of

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⁶⁸Ga(370MBq) to final pH of 4.0 and incubating at room temperature for 15min. The mixture was diluted with 5ml saline and filtered with 0.22µm membrane for quality control and imaging. Instant thin layer chromatography (ITLC) was used for ⁶⁸Ga-citrate quality control. The radiochemical purity was determined by ITLC-SG strips (Pall) with methanol and glacial acetic acid (v:v=9:1) as mobile phase.

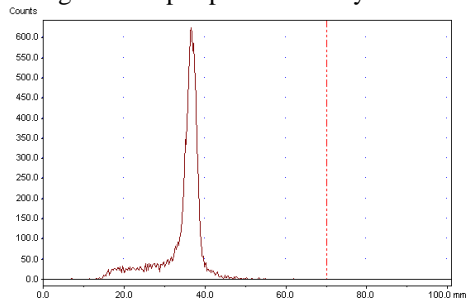
PET Imaging

⁶⁸Ga-Citrate (148 MBq) was injected intravenously in two DLBCL patients and one EATL (Type II) patient. The images were acquired 60 minutes post-injection.

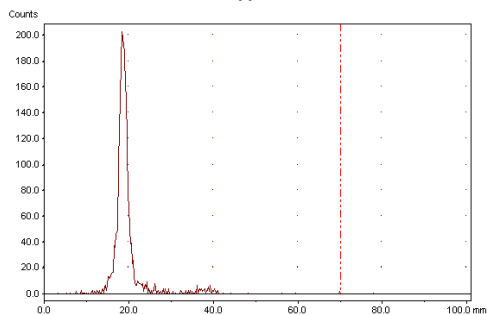
RESULTS

Quality control of ⁶⁸Ga-citrate

The radiochemical purity and yield of ⁶⁸Ga-citrate was over 99%. The Rf of ⁶⁸Ga-citrate was 1 (Fig 1A) while that of ⁶⁸Ga was 0 (Fig 1B) in mobile phase glacial acetic acid and methanol (1:9). This result indicated that the ⁶⁸Ga directly eluted from ⁶⁸Ge/⁶⁸Ga generator can be used for citrate labeling without pre-purification by ion exchange resin.



A

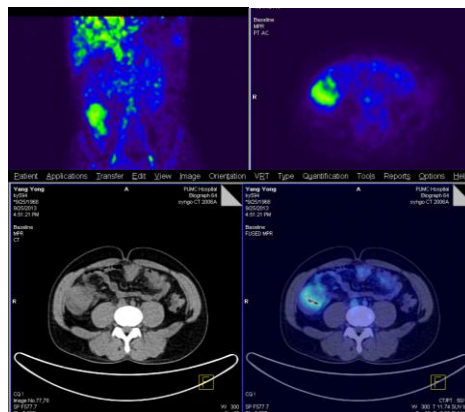


B

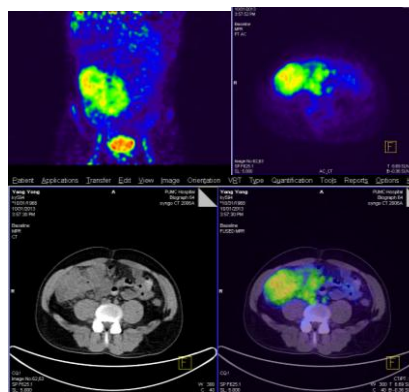
Fig 1. Radiochemical purity determination of ⁶⁸Ga-citrate by ITLC-SG with mobile phase glacial acetic acid and methanol. (A. ⁶⁸Ga-citrate, Rf=1; B. free ⁶⁸Ga, Rf=0)

⁶⁸Ga-citrate PET/CT revealed rapid progression of DLBCL

We here present a case of a 45-year-old male patient with DLBCL. He underwent ⁶⁸Ga-citrate PET/CT, which clearly showed that ileocecal region of the colon was abnormal (Figure 2 top). About one month later, the patient situation became worse, intestinal obstruction symptom appeared. ⁶⁸Ga-citrate PET/CT were repeated, expanded ⁶⁸Ga-citrate accumulation was found in the previous ileocecal region of the colon (Figure 2 bottom). The lesion was removed by surgery and the biopsy of the lesion confirmed DLBCL (Figure 3). This case indicated that ⁶⁸Ga-citrate PET/CT should be used as routine procedure for diagnosis of bowel lymphoma.



Top



Bottom

PET, Vertical	PET, sectional
CT	CT fused with sectional PET

Fig 2. ⁶⁸Ga-citrate PET imaging revealed rapid progression of DLBCL. (The top PET images were acquired 90min after intravenous 3mCi dose administration demonstrated the colon wall accumulated ⁶⁸Ga-citrate significantly higher than the background on Sep 25, 2013; simultaneous CT demonstrated that the colon wall thickened. The bottom PET image demonstrated the colon wall accumulated ⁶⁸Ga-citrate further wider than before On Oct 31, 2013; simultaneous CT demonstrated the colon wall further thickened.

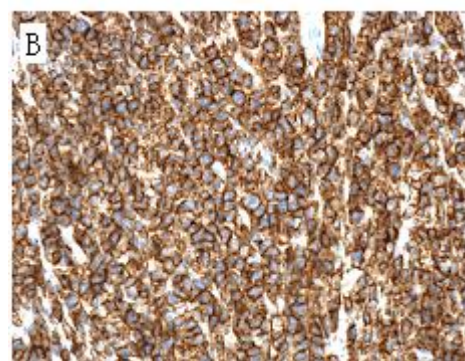
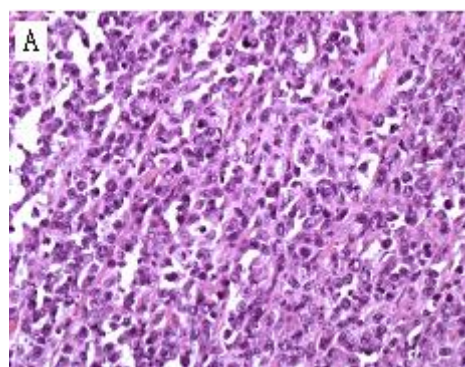


Fig 3. The Surgery biopsy of the colon lesion display large atypical lymphoid cells with prominent nucleoli (A, H&E staining). The tumor cells was positive for CD20, a pan B cell marker (B, peroxidase staining)

⁶⁸Ga-citrate revealed more lesion than ¹⁸F-FDG in DLBCL

We here present another case of a 73-year-old female patient with DLBCL. She underwent ⁶⁸Ga-citrate PET/CT and then ¹⁸F-FDG PET/CT in consecutive day. ⁶⁸Ga-citrate PET/CT revealed DLBCL lesions in omentum majus, ileocecal junction, and ileum (Figure 4 right). ¹⁸F-FDG PET/CT only revealed DLBCL lesion in omentum majus (Figure 4 left). The lesions were removed by surgery and DLBCL in ileum, ileocecal junction and omentum majus were confirmed by pathological staining.

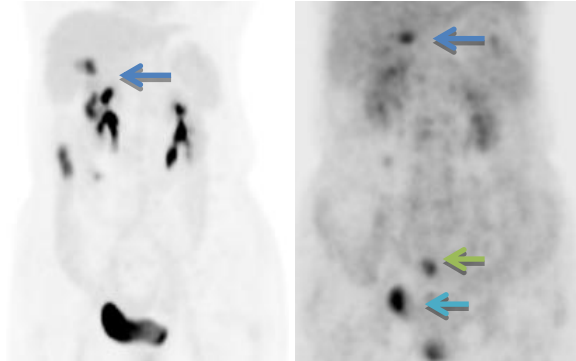


Fig 4. ⁶⁸Ga-citrate detected ileac DLBCL lesion, but ¹⁸F-FDG did not. Omentum majus (blue arrow), ileum (green arrow), ileocecal junction (brilliant blue arrow)

⁶⁸Ga-citrate imaging was negative in an EATL patient

We here present another case of an 82-year-old male patient with EATL (Type II) in ascending colon. He underwent ⁶⁸Ga-citrate PET/CT (Figure 5 right) and ¹⁸F-FDG PET/CT (Figure 5 left) in consecutive day. Although both tracers show negative accumulation in ascending colon, the biopsy demonstrated that EATL lesion was present in ascending colon.

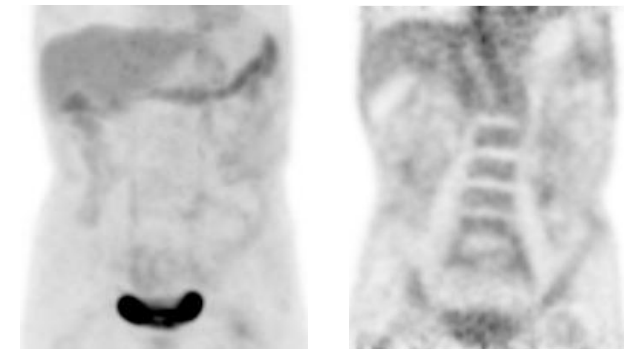


Fig 5. ⁶⁸Ga-citrate imaging was negative in one EATL (Type II) patient (right) while ¹⁸F-FDG imaging was also negative (left).

DISCUSSION

Primary colorectal lymphoma is rare gastrointestinal lymphomas (3%) and DLBCL is the most common histological subtype of colorectal lymphoma [17, 18]. EATL is also rare gastrointestinal lymphomas (9%) while Type II EATL represents only approximately 10% to 20% of all EATL [19]. It has been reported that ¹⁸F-FDG PET/CT was applicable in detecting DLBCL and EATL [20-22] but ⁶⁸Ga-citrate PET/CT was not reported in DLBCL and EATL.

The literature reported that the pre-purification of ⁶⁸Ga was needed before citrate labeling. The procedure consisted of eluting ⁶⁸Ga from generator with diluted hydrochloric acid, loading ⁶⁸Ga on cationic exchange resin, washing with low acidic acetone to remove impurities such as Ti, Zn, Fe and ⁶⁸Ge. The final purified ⁶⁸Ga was eluted with higher concentration of acidic acetone and residual acetone was evaporated with 1 min boiling [14]. Dawson firstly described colorectal lymphoma in 1961 [23], but till now the lack of specific symptoms usually lead to delayed diagnosis in large part of patients. In this study, ⁶⁸Ga eluted from generator was directly added into sodium citrate for citrate labeling. The labeling efficiency is high enough for imaging without further purification. Three abdominal lymphoma cases reported here proved that ⁶⁸Ga-citrate PET/CT is valuable for diagnosis of abdominal lymphoma. The site most commonly involved in colon lymphoma is the ileocecal region due to the proliferation of lymphoid tissue (Peyer's patches) in this area [24]. In one patient, rapid progression of DLBCL was revealed by ⁶⁸Ga-citrate PET/CT. In another DLBCL, ⁶⁸Ga-citrate revealed more lesion than ¹⁸F-FDG. The SUVmax in the lymphoma of ⁶⁸Ga-citrate is lower than that of ¹⁸F-FDG. The reason for negative imaging of ⁶⁸Ga-citrate in an EATL patient is unknown. The mechanism of ⁶⁸Ga-citrate imaging is its transferrin receptor binding. Since some lymphomas are transferrin receptor negative, the lesions displayed on ⁶⁸Ga-citrate may be false negative [20]. Early detection with ¹⁸F-FDG PET/CT, combined with ⁶⁸Ga-citrate PET/CT, may be beneficial for suspected lymphoma patients. The imaging of ¹⁸F-FDG is based on glucose utilization, so the lesion displayed on ¹⁸F-FDG image may be inflammation, which results in false positive. The balance reading between the ¹⁸F-FDG and ⁶⁸Ga-citrate image is helpful for diagnosis of suspected lymphoma patients before biopsy. However, only a few cases were presented in this study, further evaluation of ⁶⁸Ga-citrate with more cases in lymphoma imaging is needed in the future.

CONCLUSION

In summury, ⁶⁸Ga-citrate without nuclide pre-purification is prepared in our facility. ⁶⁸Ga-citrate can be used for lymphoma imaging and ⁶⁸Ga-citrate PET/CT is proved to be valuable for abdominal lymphoma imaging. Further ⁶⁸Ga-citrate PET/CT evaluation in lymphoma detection is warranted.

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REFERENCES

- [1] Ferreri A.J., "Risk of CNS dissemination in extranodal lymphomas." *Lancet Oncology*, vol. 15, 2014, pp. e159-69. W.-K. Chen, *Linear Networks and Systems* (Book style). Belmont, CA: Wadsworth, 1993, pp. 123-135.
- [2] C.Y. Lim, K.O. Ong., "Imaging of musculoskeletal lymphoma." *Cancer Imaging*, vol. 13, 2013, pp. 448-457.
- [3] G. da Cunha Santos, H.M. Ko, W.R. Geddie, S.L. Boerner, S.W. Lai, C. Have, S. Kamel-Reid, D. Bailey, "Targeted use of fluorescence in situ hybridization (FISH) in cytospin preparations: results of 298 fine needle aspirates of B-cell non-Hodgkin lymphoma." *Cancer Cytopathology*, vol. 118, 2010, pp. 250-258

- [4] R. Küppers, A. Engert, M.L. Hansmann, "Hodgkin lymphoma." *Journal Clinical Investigation*, Vol. 122, 2012, pp. 3439-3447.
- [5] T. Interimesoli, A. Rambaldi, G. Rossi, F. Delaini, C. Romani, E.M. Pogliani, C. Pagani, E. Angelucci, E. Terruzzi, A. Levis, V. Cassibba, D. Mattei, G. Gianfaldoni, A.M. Scattolin, E. Di Bona, E. Oldani, M. Parolini, N. Gökbüget, R. Bassan, "High cure rates in Burkitt lymphoma and leukemia: a Northern Italy Leukemia Group study of the German short intensive rituximab-chemotherapy program." *Haematologica*, vol. 98, 2013, pp.1718-1725.
- [6] M.Roschewski, L.M.Staudt, W.H. Wilson, "Diffuse large B-cell lymphoma-treatment approaches in the molecular era." *Nat Review Clinical Oncology*, vol. 11, 2014, pp. 12-23.
- [7] M. Mian, F. Augustin, F. Kocher, E. Gunsilius, W. Willenbacher, A. Zabernigg, G. Zangerl, H. Oexle, S. Schreieck, M. Schnallinger, M. Fiegl. "A success story: how a single targeted-therapy molecule impacted on treatment and outcome of diffuse large B-cell lymphoma." *Anticancer Research*, vol. 34, 2014, pp. 2559-2564.
- [8] K.O.Franssila, N.Jaser, A.Sivula, "Gastrointestinal non-Hodgkin's lymphoma. A population-based clinic pathological study of 111 adult cases with a follow-up of 10-15 years." *APMIS*, vol. 101, 1993, pp. 631-641.
- [9] S.Adler, K.L.Parthasarathy, S.P.Bakshi, L.Stutzman, "Gallium-67-citrate scanning for the localization and staging of lymphomas." *Journal Nuclear Medicine*, vol. 16, 1975, pp.255-260
- [10] J.A. Levi, M.J. O'Connell, W.L. Murphy, J.C. Sutherland, P.H.Wiernik, "Role of 67gallium citrate scanning in the management of non-Hodgkin's lymphoma." *Cancer*, vol. 36, 1975, pp. 1690-1741.
- [11] J. E. Seabold, M. L. Votaw, J. W. Keyes, W.D. Foley, S. Balachandran, S. P. Gill, "Gallium citrate Ga 67 scanning: clinical usefulness in lymphoma patients." *Archives Internal Medicine*, vol. 136, 1976 , pp.1370-1374
- [12] J. Okada, K. Yoshikawa, K. Imazeki, S. Minoshima, K. Uno, J. Itami, J. Kuyama, H. Maruno, N. Arimizu, "The use of FDG-PET in the detection and management of malignant lymphoma: correlation of uptake with prognosis." *Journal Nuclear Medicine*, vol. 32, 1991, pp. 686-91.
- [13] M. Gasparini, E. Bombardieri, M. Castellani, C. Tondini, L. Maffioli, L. Devizzi, P. Gerundini, "Gallium-67 scintigraphy evaluation of therapy in non-Hodgkin's lymphoma." *Journal Nuclear Medicine*, vol. 39, 1998, pp.1586-1590.
- [14] V. Kumar, D.K. Boddeti, S.G. Evans, S. Angelides. "(68)Ga-Citrate-PET for diagnostic imaging of infection in rats and for intra-abdominal infection in a patient." *Current Radiopharmaceutical*, vol. 5, 2012, pp. 71-75.
- [15] C. Nanni, C. Errani, L. Boriani, L. Fantini, V. Ambrosini, S. Boschi, D. Rubello, C. Pettinato, M. Mercuri, A. Gasbarrini, S. Fanti, "68Ga-citrate PET/CT for evaluating patients with infections of the bone: preliminary results." *Journal Nuclear Medicine*, vol. 51, 2010, pp.1932-1936.
- [16] A. Rizzello, D. Di Pierro, F. Lodi, S. Trespidi, G. Cicoria, D. Pancaldi, C. Nanni, M. Marengo, M.C. Marzola, A. Al-Nahhas, D. Rubello, S. Boschi, "Synthesis and quality control of 68Ga citrate for routine clinical PET." *Nuclear Medicine Communication*, vol. 30, 2009, pp. 542-545.
- [17] M.T. Wong, K.W. Eu, "Primary colorectal lymphomas." *Colorectal Disease*, vol. 8, 2006, pp. 586-591.
- [18] Q.H. Gonzalez, M.J. Heslin, A. Dávila-Cervantes, "Primary colonic lymphoma." *American Surgery*, vol. 74, 2008, pp. 214-216
- [19] A. J. Ferreri, P. L. Zinzani, S. Govi, "Enteropathy-associated T-cell lymphoma." *Critical Reviews in Oncology Hematology*, vol. 79, 2011, 84-90.
- [20] T. Tsujikawa, H. Okazawa, T. Tsuchida, Y. Demura, Y. Imamura, Y. Fujibayashi, "A 18F-FDG-positive, 67Ga-negative, and transferrin receptor expression-negative patient with diffuse large B-cell lymphoma." *Annals Nuclear Medicine*, vol. 21, 2007, pp. 375-378.
- [21] M. Hadithi, M. Mallant, J. Oudejans, J. H. van Waesberghe, C. J. Mulder, E. F. Comans, "18F-FDG PET versus CT for the detection of enteropathy-associated T-cell lymphoma in refractory celiac disease." *Journal Nuclear Medicine*, vol. 47, 2006, pp. 1622-1627.
- [22] C. Casulo, H. Schöder, J. Feeney, R. Lim, J. Maragulia, A. D. Zelenetz, S. Horwitz, "18F-fluorodeoxyglucose positron emission tomography in the staging and prognosis of T cell lymphoma." *Leukemia Lymphoma*, vol. 54, 2013, pp. 2163-2167.
- [23] I. M. Dawson, J. S. Cornes, B. C. Morson, "Primary malignant lymphoid tumours of the intestinal tract. Report of 37 cases with a study of factors influencing prognosis." *British Journal Surgery*, vol. 49, 1961, pp. 80-89.
- [24] M. T. Wong, K. W. Eu, "Primary colorectal lymphomas." *Colorectal Disease*, vol. 8, 2006, pp. 586-591.